

The Crystal and Molecular Structure of the Dimeric Copper(II) Chelate of Glycyl-L-leucyl-L-tyrosine*

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The crystal structure of the copper(II) chelate of glycyl-L-leucyl-L-tyrosine, prepared at neutral pH, has been determined and refined by three-dimensional least-squares techniques using 2043 counter data. The crystals are orthorhombic, space group $P2_12_12_1$, with $a=9.316$, $b=25.76$ and $c=21.05$ Å. The unit-cell contains four dimeric units of $\text{Cu}_2(\text{glt})_2$, 32 molecules of water and four molecules of diethyl ether; thus, there are two independent peptide units in the asymmetric unit. The final R value is 0.103. The packing of the peptides is very similar to an anti-parallel pleated sheet structure. It is concluded that the conformation of the peptide is only affected slightly by the chelation. There seems to exist a weak interaction of the copper ions with the π -system of the aromatic rings of the tyrosine groups and the suggestion is made that this interaction correlates with the oxidase activity of certain copper-containing proteins and enzymes.

Introduction

The structure of the copper (II) chelate of glycyl-L-leucyl-L-tyrosine (glt) was determined as one in a series of peptide chelate studies to investigate the influence of metal ions on the conformation of peptides and to study the coordination of metal ions in these complexes as well as the hydrogen bonding which occurs. The peptide seemed particularly suitable because both residues contain side chains and one – the tyrosine residue – has an extra functional group. Furthermore the preliminary work indicated the occurrence of two peptides and two copper ions in the asymmetric unit, allowing comparisons of two independent molecules. A note on the unusual coordination geometry observed in this structure has been published (Van der Helm & Franks, 1968).

Experimental results

The method which was most consistent in yielding crystals of the copper complex of glt proceeded by reacting equimolar quantities of CuSO_4 , $\text{Ba}(\text{OH})_2$, both in aqueous solution, and powdered glt. The BaSO_4 was removed by filtration and the resulting chelate solution was approximately 0.055 M in peptide. The aqueous solution (pH 6–7) was equilibrated with diethyl ether and suitable crystals appeared on standing overnight. The crystals of the complex were well-

formed blue plates which decomposed within one minute on exposure to the atmosphere.

For the X-ray experiments a crystal was transferred in its mother liquor to a thin-walled (0.01 mm) glass capillary. The mother liquor in the capillary was layered with ether. The capillary was then sealed with a small flame during which it was found useful to immerse the capillary in ice water. The systematic absences, $h00(h \neq 2n)$, $0k0(k \neq 2n)$ and $00l(l \neq 2n)$, showed the space group to be $P2_12_12_1$. The 2θ -values of 49 reflections were measured at room temperature (23°C) and used in a least-squares calculation to obtain the cell dimensions which are $a=9.316 \pm 0.002$, $b=25.76 \pm 0.02$ and $c=21.05 \pm 0.01$ Å. The density of the crystal could not be measured owing to the instability of the compound. The cell dimensions together with a likely value of the density indicated the contents of the asymmetric unit to be $\text{Cu}_2(\text{glt})_2 \cdot (8-12)\text{H}_2\text{O}$. It was not until the structure was fully determined that the actual contents of the asymmetric unit were found to be $\text{Cu}_2(\text{C}_{17}\text{H}_{23}\text{N}_3\text{O}_5)_2 \cdot 8\text{H}_2\text{O} \cdot \text{C}_4\text{H}_{10}\text{O}$, with a formula weight of 1044.1. The calculated density is 1.37 $\text{g}\cdot\text{cm}^{-3}$ for $Z=4$.

The integrated intensities were measured using Ni-filtered $\text{Cu } K\alpha$ radiation and the θ - 2θ scan technique on a General Electric XRD-5 diffractometer equipped with a single-crystal orienter and a scintillation counter. The background counts were fairly high and few reflections beyond 2θ of 90° had an observable intensity. Of the 2252 independent reflections with a 2θ value below 90°, 2044 were observed and used for the subsequent structure determination. Lorentz, polarization and absorption corrections were applied to the data. The linear absorption factor assumed was 1.80 cm^{-1} , although on the basis of the composition indicated by the final results of the structure determination a value of 17.0 cm^{-1} was calculated. The presence of the

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Table 1. *Parameters of all carbon, nitrogen and oxygen atoms*

Calculated standard deviations for the last digit are given in parentheses.
 Fractional molecule of water, *W*(1), was located from the last difference Fourier synthesis.

	<i>x</i>	<i>y</i>	<i>z</i>	<i>B</i>
Molecule A				
N(1)	0.525 (2)	-0.0409 (8)	0.0384 (10)	6.0 (5)
C(1)	0.634 (2)	-0.0280 (8)	0.0847 (9)	3.0 (4)
C(2)	0.566 (2)	-0.0133 (7)	0.1454 (9)	2.2 (4)
O(1)	0.430 (1)	-0.0143 (4)	0.1505 (6)	2.5 (3)
N(2)	0.651 (2)	-0.0004 (6)	0.1918 (7)	3.0 (4)
C(3)	0.598 (2)	0.0099 (7)	0.2598 (9)	2.7 (4)
C(4)	0.663 (2)	0.0611 (7)	0.2776 (9)	2.3 (4)
O(2)	0.791 (2)	0.0714 (6)	0.2632 (7)	4.2 (3)
N(3)	0.583 (2)	0.0934 (6)	0.3087 (7)	3.1 (3)
C(5)	0.648 (2)	0.1423 (9)	0.3214 (10)	4.1 (5)
C(6)	0.585 (3)	0.1627 (10)	0.3861 (10)	5.4 (6)
O(3)	0.470 (2)	0.1442 (6)	0.4059 (7)	4.6 (4)
O(4)	0.653 (2)	0.2013 (7)	0.4112 (9)	7.6 (5)
C(7)	0.620 (3)	0.1859 (10)	0.2676 (10)	6.0 (6)
C(8)	0.460 (3)	0.1923 (10)	0.2557 (10)	6.0 (7)
C(9)	0.382 (3)	0.1611 (10)	0.2212 (10)	5.2 (6)
C(10)	0.229 (3)	0.1669 (10)	0.2125 (10)	6.2 (7)
C(11)	0.166 (3)	0.2039 (10)	0.2557 (10)	7.0 (7)
C(12)	0.241 (3)	0.2371 (10)	0.2837 (10)	7.5 (8)
C(13)	0.381 (3)	0.2313 (10)	0.2940 (10)	7.0 (7)
O(5)	0.013 (2)	0.2047 (8)	0.2499 (10)	8.2 (5)
C(14)	0.649 (2)	-0.0343 (8)	0.3033 (9)	3.4 (5)
C(15)	0.611 (3)	-0.0900 (10)	0.2804 (10)	4.9 (5)
C(16)	0.448 (4)	-0.0944 (10)	0.2847 (20)	9.5 (10)
C(17)	0.683 (4)	-0.1284 (10)	0.3257 (20)	8.6 (9)
Molecule B				
N(1)	0.193 (2)	0.1112 (6)	0.3785 (8)	3.7 (4)
C(1)	0.080 (2)	0.0862 (8)	0.3411 (9)	3.1 (4)
C(2)	0.151 (2)	0.0491 (7)	0.2917 (9)	2.9 (4)
O(1)	0.285 (1)	0.0420 (5)	0.2950 (6)	3.3 (3)
N(2)	0.069 (2)	0.0264 (6)	0.2533 (7)	2.9 (4)
C(3)	0.123 (2)	-0.0129 (7)	0.2052 (8)	1.7 (4)
C(4)	0.057 (2)	0.0029 (8)	0.1412 (10)	3.1 (5)
O(2)	-0.076 (1)	0.0169 (5)	0.1398 (6)	3.1 (3)
N(3)	0.139 (2)	-0.0019 (6)	0.0901 (7)	3.3 (4)
C(5)	0.066 (2)	0.0115 (8)	0.0303 (10)	3.5 (5)
C(6)	0.122 (2)	-0.0278 (8)	-0.0181 (9)	2.8 (4)
O(3)	0.243 (2)	-0.0506 (6)	-0.0083 (7)	4.1 (3)
O(4)	0.059 (1)	-0.0308 (6)	-0.0706 (7)	4.4 (3)
C(7)	0.092 (3)	0.0661 (9)	0.0087 (10)	4.8 (6)
C(8)	0.259 (3)	0.0781 (10)	-0.0014 (10)	4.7 (6)
C(9)	0.336 (3)	0.0924 (10)	0.0448 (10)	6.2 (6)
C(10)	0.498 (3)	0.0998 (10)	0.0369 (10)	6.7 (7)
C(11)	0.549 (3)	0.0854 (10)	-0.0211 (10)	6.1 (7)
C(12)	0.467 (3)	0.0700 (10)	-0.0642 (10)	7.1 (7)
C(13)	0.320 (3)	0.0634 (10)	-0.0609 (10)	7.1 (7)
O(5)	0.697 (3)	0.0891 (10)	-0.0256 (10)	11.3 (7)
C(14)	0.060 (3)	-0.0650 (10)	0.2309 (10)	5.3 (6)
C(15)	0.092 (4)	-0.1104 (10)	0.1863 (10)	7.8 (8)
C(16)	-0.031 (4)	-0.1197 (10)	0.1370 (20)	10.0 (10)
C(17)	0.094 (5)	-0.1638 (20)	0.2347 (20)	11.7 (10)
Water molecules				
<i>W</i> (1)	-0.417	-0.159	0.037	19.0
<i>W</i> (2)	0.176 (3)	-0.1875 (10)	0.0034 (10)	13.0 (8)
<i>W</i> (3)	-0.140 (4)	-0.2342 (10)	0.0413 (10)	16.2 (10)
<i>W</i> (4)	-0.180 (3)	0.1564 (10)	0.0362 (10)	13.0 (8)
<i>W</i> (5)	-0.261 (3)	-0.0382 (10)	-0.0612 (10)	12.3 (8)
<i>W</i> (6)	0.356 (4)	-0.1265 (10)	0.0918 (20)	17.9 (10)
<i>W</i> (7)	0.070 (3)	-0.2754 (9)	0.1182 (10)	11.2 (7)
<i>W</i> (8)	-0.110 (3)	0.1338 (10)	0.1602 (10)	11.9 (7)
Ether molecule				
C(18)	0.371 (6)	-0.2406 (20)	0.3782 (30)	17.4 (20)
C(19)	0.365 (6)	-0.2132 (20)	0.4342 (30)	17.9 (20)
O(9)	0.348 (4)	-0.2504 (10)	0.4838 (20)	15.4 (10)
C(20)	0.363 (7)	-0.2327 (20)	0.5393 (30)	19.0 (20)
C(21)	0.397 (7)	-0.2677 (20)	0.5881 (30)	17.7 (20)

capillary and the liquid were neglected in the absorption calculations.

Structure determination

The positions of the two copper atoms were unambiguously located from a sharpened Patterson synthesis. In order to locate more atoms an eightfold superposition map and a Fourier synthesis phased on the copper atom locations were used simultaneously. It was possible to locate the benzene groups and peptide chains. Three side-chain atoms, *i.e.* two from isopropyl groups and one from a benzene ring, were not found in either map. The peaks of some water molecules were present in both maps. A structure factor calculation was made using the peptide atoms which were located and the two copper atoms. The resulting *R* index ($= \sum ||kF_o| - |F_c|| / \sum |kF_o|$) was 0.32.

The trial structure was refined by means of block-diagonal least-squares computations (4×4 and 9×9) using anisotropic temperature factors for the copper atoms and isotropic ones for the light atoms. When the *R* value was 0.17 a difference Fourier synthesis was calculated. It showed the remaining three side-chain carbon atoms and seven water molecules. These atoms were included in subsequent least-squares cycles.

This first difference Fourier synthesis also showed four peaks which were clustered together at distances smaller than hydrogen bond lengths. In a second difference synthesis, calculated when the *R* index was 0.125, a fifth peak appeared in close proximity to the four previously observed. The five peaks formed an extended chain with distances of 1.34, 1.50, 1.38 and 1.34 Å. The peak in the middle had a height of $2.1 \text{ e.}\text{\AA}^{-3}$ where the other four ranged between 1.26–1.39 $\text{e.}\text{\AA}^{-3}$. In addition the peak in the middle was located at a hydrogen bond distance from a water molecule where the distances of the remaining peaks to other atoms in the structure were all larger than 3.5 Å. The group of peaks was therefore identified as diethyl ether, thus explaining the necessity of this solvent for crystallization. All atoms so far located were included in a final series of least-squares calculations. The refinement was terminated when the parameter shifts were smaller than $\frac{1}{3}$ of the calculated standard deviations. The most pronounced features in the subsequent difference synthesis were two peaks with heights of 1.0 and 0.6 $\text{e.}\text{\AA}^{-3}$, which were close together and at normal hydrogen bond distances from other atoms in the structure. The two peaks had already been observed in the previous difference Fourier synthesis. The peaks were assigned as a disordered molecule of

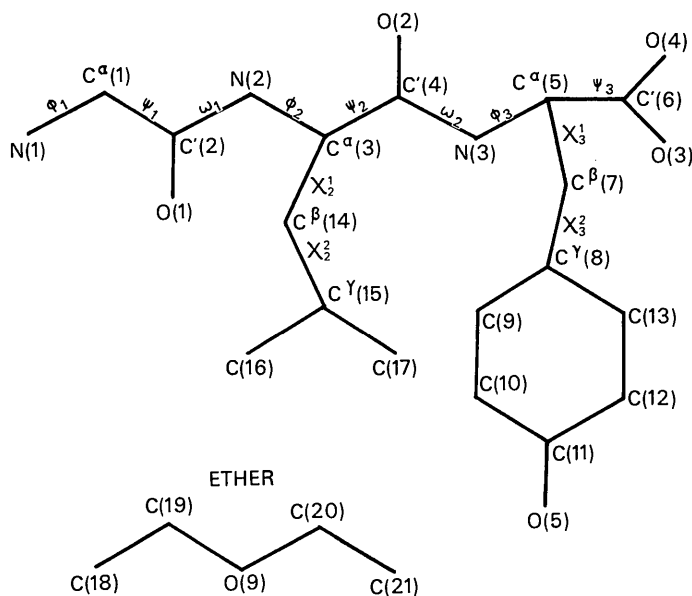


Fig. 1. Numbering of atoms and notation for conformational angles.

Table 2. Parameters for the copper atoms

The anisotropic temperature factors are expressed in the form:

$$\exp [-(b_{11}h^2 + b_{22}k^2 + b_{33}l^2 + b_{23}kl + b_{13}hl + b_{12}hk)].$$

	<i>x</i>	<i>y</i>	<i>z</i>	<i>b</i> ₁₁	<i>b</i> ₂₂	<i>b</i> ₃₃	<i>b</i> ₂₃	<i>b</i> ₁₃	<i>b</i> ₁₂
Cu(1)	0.3268 (3)	-0.0285 (1)	0.0721 (1)	0.0087 (4)	0.0031 (1)	0.0020 (1)	0.0008 (1)	0.0000 (3)	0.0006 (3)
Cu(2)	0.3935 (3)	0.0911 (1)	0.3504 (1)	0.0094 (3)	0.0021 (1)	0.0021 (1)	0.0010 (1)	0.0002 (3)	0.0003 (3)

Table 3 (cont.)

Table with multiple columns containing numerical data and headers such as 'K FO FC ALPHA', 'K FO FC ALPHA', etc., organized in a grid-like structure.

a common scale with the standard reflection while those with low intensities were affected by adverse counting statistics. The final positional and thermal parameters and their estimated standard deviations, as calculated from the inverse of the least-squares matrix, are given in Tables 1 and 2. No attempt was

made to locate the hydrogen atoms in view of the stated purposes of the structure determination, the high temperature factors observed and the large size of the observed amplitudes used in the refinement with the 20. When the 311 unobserved reflections within the 2θ

limit of 90° are included, the *R* value is 0.108. A list of the observed and calculated structure factors is given in Table 3. The absorption corrections were calculated on an IBM 1410 computer with a program written by P. J. Schapiro. All other computations were carried out on an IBM 360-40. The structure factor-least squares and Fourier programs used were written by Ahmed (1966*a,b*). Table 3 was prepared with a program written by Pippy (1967). The atomic scattering factors used for Cu²⁺, O, N and C were those listed in *International Tables for X-ray Crystallography* (1962).

Description and discussion of the structure

General, chelation sites and metal surrounding

A schematic drawing of a peptide and ether molecule, specifying the atom numbering used, is shown in Fig. 1. Fig. 2 shows a projection of a part of the structure down the *b* axis.

The atomic positions of Table 1 correspond to the atoms of peptide chains *A* and *B*. Some of the carbon atoms in these peptide units are identified in Fig. 2 for clarification. Some of the water molecules are drawn but none of the ether molecules. The reason for the limitations is the length of the projection axis, 25.76 Å. This particular projection, however, shows most clearly the basic building blocks of the structure which are copper-peptide dimers. One dimer is identified in Fig. 2 by labels *A* and *B*, for the peptides, Cu(1) and Cu(2). The modes of chelation of the two peptide groups to the metal ions are quite similar. Two

five-membered chelate rings are formed by means of the terminal nitrogen atoms of the glycyl residues, N(1), and the carbonyl oxygen atoms of the glycyl-leucyl peptide groups O(1) of molecules *A* and *B* with Cu(1) and Cu(2) respectively. Two other five-membered chelate rings are formed by the deprotonated amide nitrogen atom of the leucyl-tyrosine peptide groups, N(3), and an oxygen atom of the terminal acid groups of *A* and *B* with Cu(2) and Cu(1) respectively. Another proof that these particular peptide nitrogen atoms are deprotonated in this complex, formed at pH 6-7, is the trigonal surrounding of both N(3) atoms by Cu, C(4) and C(5). The sum of the bond angles is 360° for peptide N(3*A*) and 359° for N(3*B*) (see Tables 4 and 7). It should be noted that both peptides are doubly deprotonated on the acid end of the molecule.

The five-membered chelate rings form the bases for the square-pyramidal surroundings of Cu(1) and Cu(2). The tops of the pyramids are W(6) for Cu(1) and O(4*B*) (*e*) for Cu(2). These fifth bonds are somewhat weaker than those in the basal plane (Van der Helm & Franks, 1968). The atoms forming the bases of the pyramids are tetrahedrally distorted (planes 1 and 2, Table 4), while the copper ions are displaced from the base plane toward the top of the pyramid. Angles and distances in the coordination of the two independent copper ions are given in Table 5. None of the four five-membered chelate rings are planar (planes 3, 4, 5 and 6, Table 4). The rings involving the free acid group are in the half-chair conformation with the peptide nitrogen and copper ion on opposite sides of the least-squares planes through the acid groups. The chelate

Table 4. *Least-squares planes*

The equations of the planes are expressed in the form:

$$Ax + By + Cz = D$$

where *x*, *y* and *z* are fractional coordinates and *D* is the distance from the origin in Å. The method of Schomaker, Waser, Marsh & Bergman (1959) was used to calculate the least-squares planes.

1	N(1 <i>A</i>), O(1 <i>A</i>), O(3 <i>B</i>) N(3 <i>B</i>)	1.481	23.887	-7.140	-0.631
2	N(1 <i>B</i>), O(1 <i>B</i>), O(3 <i>A</i>), N(3 <i>A</i>)	1.731	-18.412	14.193	3.789
3	C(1 <i>A</i>), C(2 <i>A</i>), O(1 <i>A</i>), N(2 <i>A</i>)	0.374	-24.573	6.269	1.454
4	C(5 <i>A</i>), C(6 <i>A</i>), O(3 <i>A</i>), O(4 <i>A</i>)	5.017	-17.121	10.902	4.327
5	C(1 <i>B</i>), C(2 <i>B</i>), O(1 <i>B</i>), N(2 <i>B</i>)	1.184	19.556	-13.438	-2.798
6	C(5 <i>B</i>), C(6 <i>B</i>), O(3 <i>B</i>), O(4 <i>B</i>)	4.929	19.242	-8.476	0.276
7	C(3 <i>A</i>), C(4 <i>A</i>), O(2 <i>A</i>), N(3 <i>A</i>)	2.913	-10.135	18.195	6.368
8	C(3 <i>B</i>), C(4 <i>B</i>), O(2 <i>B</i>), N(3 <i>B</i>)	2.768	24.374	2.720	0.590

	Δ (1)	Δ (2)	Δ (3)	Δ (5)
N(1)	+0.16 Å	-0.13 Å	-0.00 Å	-0.01 Å
O(1)	-0.15	+0.12	-0.00	+0.02
O(3)	-0.16	+0.13	+0.00	-0.01
N(3)	+0.15	-0.12	+0.00	-0.01
Cu	-0.08	+0.19	-0.01	+0.12
			-0.18	+0.34
			+0.16	-0.07

	Δ (4)	Δ (6)	Δ (7)	Δ (8)
C(5)	-0.01	+0.02	0.00	-0.01
C(6)	+0.03	-0.06	-0.00	+0.02
O(4)	-0.01	+0.02	0.00	-0.01
O(3)	-0.01	+0.02	0.00	-0.01
N(3)	+0.37	-0.39	-0.98	+0.93
Cu	-0.09	+0.18	-0.08	-0.04

rings involving the free amino group are in the envelope conformation.

It was pointed out (Van der Helm & Franks, 1968) that close approaches (between 3.17 and 3.34 Å) (Table 5) occur between the copper atoms and C(8) and C(9) of both tyrosine groups. A similar and even closer approach has now been found in the structure of bis(L-tyrosinato)copper(II) (Tatsch & Van der Helm, 1969). In all three cases the side chains of L-tyrosine fold back so that they are situated below the basal planes of the square pyramidal copper coordination (Fig. 2; Van der Helm & Franks, 1968: Fig. 1). This is a result of the fact that the $C^\beta-C^\gamma$ [*i.e.* C(7)–C(8)] bonds are skewed with respect to both $N-C^\alpha$ [N(3)–C(5)] and $C^\alpha-C'$ [C(5)–C(6)] bonds. For an L-amino acid that means that the $N-C^\alpha-C^\beta-C^\gamma$ and $C'-C^\alpha-C^\beta-C^\gamma$ configurational angles have to be approximately 60° and 300°, respectively. The fact, however, that both these angles are skewed does not necessarily imply that close approaches must occur between the metal atom and the ring in the side chain. For instance in the structure of (β -alanyl-L-histidinato)copper(II) dihydrate (Freeman & Szymanski, 1967), the $C^\beta-C^\gamma$ bond of the histidine residue is skewed with respect to the $N-C^\alpha$ and $C^\alpha-C'$ bonds, but the closest metal imidazole distance is 3.52 Å. It is therefore believed that an interaction exists between the Cu^{2+} ions and the aromatic rings of the tyrosine groups both in the present structure as in the bis(L-tyrosinato)copper(II) structure. The weakness of the interaction allows for rapid kinetics of reaction. Both the enzymes tyrosinase and laccase, and the protein ceruloplasmin contain copper and have oxygenase activity. It is interesting to note that Levine & Peisach (1962) suggested, on the basis of chemical data, that the enzyme binding in ceruloplasmin to substrate was not through the amine or phenolic group but instead directly to the π -electrons of the aromatic ring. Broman, Malmström, Aasa & Vänngård (1963) hypothesized that the Cu^+ ion, which is also present in ceruloplasmin, partakes in substrate binding by interaction with the π -system of the substrate. The present suggestion, however, is the involvement of Cu^{2+} , rather than Cu^+ , in this type of binding, or interaction, with the substrate. This interaction, subsequently, allows an electronic reduction-oxidation reaction of the Cu^{2+} ion with the phenolic group.

Table 5. *Angles and distances in the metal coordination*

The literature values are from Freeman (1967). Standard deviations for the last digit are given in parentheses. O(*t*) is W(6) and O(4B) for the Cu(1) and Cu(2) coordination respectively.

	Cu(1)	Cu(2)	Literature
Cu–N(1)	2.00 (2) Å	2.02 (2) Å	2.00 (1) Å
Cu–O(1)	1.95 (2)	2.00 (2)	1.99 (1)
Cu–N(3)	1.92 (2)	1.98 (2)	1.92 (1)
Cu–O(3)	1.95 (2)	1.94 (2)	1.98 (1)
Cu–O(<i>t</i>)	2.57 (2)	2.32 (2)	
Cu–C(8)	3.21 (3)	3.34 (3)	
Cu–C(9)	3.17 (3)	3.27 (3)	

Table 5 (*cont.*)

	Cu(1)	Cu(2)	Literature
O(1)–Cu–N(1)	83 (1)°	82 (1)°	
O(3)–Cu–N(3)	85 (1)	85 (1)	
O(3)–Cu–N(1)	91 (1)	89 (1)	
O(1)–Cu–N(3)	103 (1)	102 (1)	
O(<i>t</i>)–Cu–O(1)	90 (1)	95 (1)	
O(<i>t</i>)–Cu–N(1)	79 (1)	98 (1)	
O(<i>t</i>)–Cu–N(3)	114 (1)	100 (1)	
O(<i>t</i>)–Cu–O(3)	84 (1)	88 (1)	
Cu–N(1)–C(1)	112 (1)°	113 (1)°	111 (1)°
Cu–O(1)–C(2)	115 (1)	116 (1)	113
Cu–O(3)–C(6)	111 (1)	113 (2)	115 (1)
Cu–N(3)–C(5)	109 (1)	109 (1)	116 (1)
Cu–N(3)–C(4)	136 (1)	136 (1)	120 (1)
O(1A)–N(3B)	3.02 (3) Å	O(3B)–N(1A)	2.81 (3) Å
O(1B)–N(3A)	3.09 (3)	O(3A)–N(1B)	2.78 (3)

Some of the bond angles involving the chelate rings are shown in Table 5 and compared with literature values. The significant differences are in the angles around the N(3) atom. The Cu–N(3)–C(4) angles deviate 16° from the average values (for 5 angles) reported previously (Freeman, 1967). Both in the present structure and in those to which the bond angles are compared, the hydrogen atom on the amide nitrogen atom has been ionized. The important difference between this structure and those with which it is compared, *i.e.* diaquoglycylglycinatocopper(II) hydrate (Freeman, 1967), sodiumglycylglycylglycinato cuprate(II) (Freeman, Schoone, & Sime, 1965) and disodium glycylglycylglycylglycinato cuprate(II) decahydrate (Freeman, & Taylor, 1965), is the fact that more than one chelate ring is formed between the peptide and one particular metal ion in the latter three structures, compared with only one in the present structure. This same feature is the probable cause for the observed differences, from the literature values, of the $C'-N-C^\alpha$ angles. Freeman (1967) lists an average value of 123° for this angle when the N atom is a part of a chelate ring, which is the same as observed in free peptides (Marsh & Donohue, 1967). In the present structure, however, the $C'-N-C^\alpha$ angles are 114 and 115° for the peptide nitrogen atoms involved in chelate rings, but 123° (twice) for those which are not a part of a chelate ring (Table 7). It is therefore suggested that when a peptide forms only one chelate ring with a particular metal ion the following angles can be expected around the deprotonated nitrogen atom: Cu–N– C^α , 109°, Cu–N– C' , 136° and $C'-N-C^\alpha$, 115°, rather than 116, 120 and 123° respectively.

Bond distances and bond angles

The bond angles and bond distances in the backbone of the peptides, the phenyl and isopropyl groups, and the ether molecule, are shown in Tables 6, 7 and Fig. 3 respectively. In general these values are close to those expected. There are some relatively large deviations in the side groups, but the temperature movement is

large for those atoms (Table 1) and the estimated standard deviations are therefore rather large. The bond distances in the backbone of the peptides are similar for the same bonds in the two peptides. The average bond distances compare favorably with those of copper complexes of amino acids and peptides summarized by Freeman (1967). There is one notable exception, namely the bond distances N(2)–C(3) which are 1.54 and 1.52 Å and deviate by 4σ and 3σ from the literature value of 1.455 (Marsh & Donohue, 1967) for a free peptide or 1.46 Å (Freeman, 1967) for a chelated peptide. The other N–C $^{\alpha}$ distances in the present structure are normal.

Table 6. *Bond distances in side-chains and ether molecule*

Standard deviations for the last digit are given in parentheses.

C(5)–C(7)	1.62 (3) Å	1.50 (3) Å
C(7)–C(8)	1.52 (4)	1.60 (4)
C(8)–C(9)	1.30 (4)	1.26 (4)
C(9)–C(10)	1.44 (4)	1.53 (4)
C(10)–C(11)	1.44 (4)	1.36 (4)
C(11)–C(12)	1.25 (4)	1.25 (4)
C(12)–C(13)	1.33 (4)	1.39 (4)
C(13)–C(8)	1.48 (4)	1.43 (4)
C(11)–O(5)	1.43 (4)	1.39 (4)
C(3)–C(14)	1.53 (3)	1.56 (3)
C(14)–C(15)	1.56 (3)	1.53 (4)
C(15)–C(16)	1.52 (4)	1.56 (5)
C(15)–C(17)	1.53 (4)	1.71 (5)
C(18)–C(19)	1.38 (8)	
C(19)–O(9)	1.42 (7)	
O(9)–C(20)	1.26 (6)	
C(20)–C(21)	1.40 (8)	

Table 7. *Bond angles*

Standard deviations for the last digit are given in parentheses.

Angle	Molecule A	Molecule B
N(1)–C(1)–C(2)	110 (2)°	109 (2)°
C(1)–C(2)–O(1)	120 (2)	118 (2)
C(1)–C(2)–N(2)	117 (2)	117 (2)
O(1)–C(2)–N(2)	123 (2)	125 (2)
C(2)–N(2)–C(3)	123 (2)	123 (2)
N(2)–C(3)–C(4)	105 (2)	106 (1)
N(2)–C(3)–C(14)	109 (2)	103 (1)
C(14)–C(3)–C(4)	112 (2)	112 (2)
C(3)–C(4)–O(2)	121 (2)	119 (2)
C(3)–C(4)–N(3)	117 (2)	117 (2)
O(2)–C(4)–N(3)	122 (2)	124 (2)
C(4)–N(3)–C(5)	115 (2)	114 (2)
N(3)–C(5)–C(6)	107 (2)	105 (2)
N(3)–C(5)–C(7)	115 (2)	114 (2)
C(6)–C(5)–C(7)	108 (2)	112 (2)
C(5)–C(6)–O(3)	119 (2)	119 (2)
C(5)–C(6)–O(4)	115 (2)	118 (2)
O(3)–C(6)–O(4)	126 (2)	121 (2)
C(5)–C(7)–C(8)	110 (2)	112 (2)
C(7)–C(8)–C(9)	125 (2)	120 (2)
C(7)–C(8)–C(13)	118 (2)	117 (2)
C(9)–C(8)–C(13)	116 (3)	122 (3)
C(8)–C(9)–C(10)	124 (2)	121 (2)
C(9)–C(10)–C(11)	113 (2)	114 (2)
C(10)–C(11)–C(12)	121 (3)	122 (3)
C(10)–C(11)–O(5)	111 (2)	113 (2)
C(12)–C(11)–O(5)	126 (3)	125 (3)
C(11)–C(12)–C(13)	123 (3)	127 (3)
C(8)–C(13)–C(12)	118 (3)	114 (3)
C(3)–C(14)–C(15)	115 (2)	112 (2)
C(14)–C(15)–C(16)	106 (2)	112 (3)
C(14)–C(15)–C(17)	108 (2)	105 (2)
C(16)–C(15)–C(17)	111 (2)	106 (3)
C(18)–C(19)–O(9)	107 (4)	
C(19)–O(9)–C(20)	115 (4)	
O(9)–C(20)–C(21)	118 (5)	

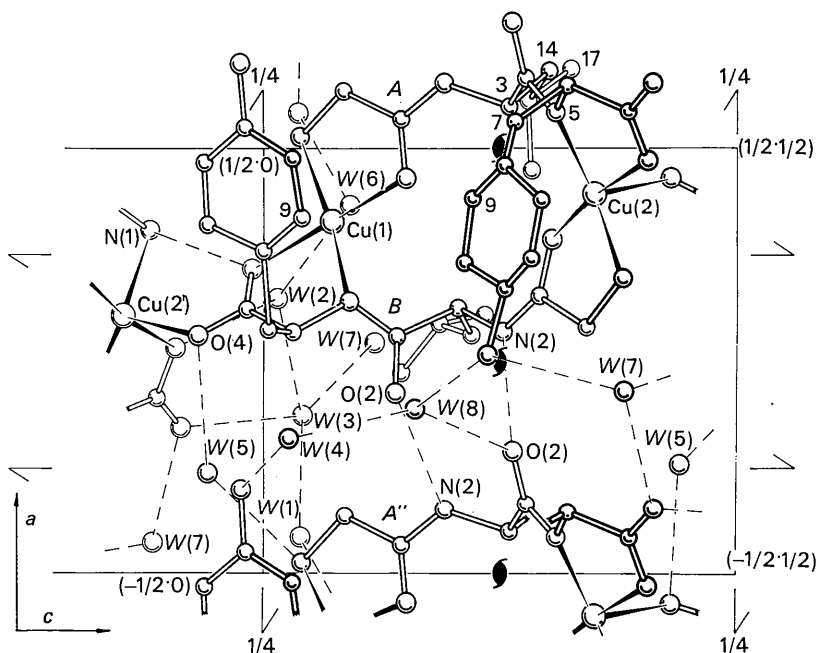


Fig. 2. Projection of a part of the structure down the *b* axis. The dashed lines are hydrogen bonds. The numbers indicate carbon atoms.

Conformational angles and sheet structure

Least-squares planes were calculated through the four atoms of each of four peptide groups $C^\alpha C' ON$ (planes 3, 5, 7 and 8, Table 4) and the acid groups (planes 4 and 6, Table 4). These groups are planar. Due to the partial double-bond character of the $C'-N$ bond the $C^{\alpha+1}$ atom is supposed to be in the plane of $C^\alpha C' ON$. In the present structure the largest deviation occurs for C(3A) (plane 3), 0.16 Å. If least-squares plane 3 is recalculated including the parameters of C(3A), the deviations from the plane are 0.04 Å for C(3A), while the largest deviation is for N(2A) 0.06 Å.

An alternative way to describe the peptide backbone is by the use of conformational angles. The conventions and nomenclature adopted for this purpose (Edsall, Flory, Kendrew, Liguori, Némethy, Ramachandran & Scheraga, 1966) will be used. The angles used are shown in Fig. 1. The calculated values of these angles in the backbone of the peptides for the present structure are given in Table 8. The standard deviation of all conformational angles is estimated to be 2° . The similarity of the φ , ψ and ω angles for peptides *A* and *B* illustrates that both have the same conformation.

Table 8. Conformational angles

The ψ_3^1 and ψ_3^2 are the conformational angles involving O(3) and O(4) respectively. The φ_1 is the rotational angle of C(1)–C(2) with respect to Cu–N(1). The hydrogen atoms are not located in the structure and ω and φ angles are therefore calculated on the assumption that the bonds $C'-N$, $N-H$ and $N-C^\alpha$ form a planar set. The columns APPS and PPS give the conformational angles for the antiparallel pleated sheet and parallel pleat structure respectively which were originally proposed by Pauling & Corey (1953). The models proposed by Pauling & Corey were built from D-amino acids. The values given in the Table are for the same models but now built from L-amino acids. The original use of D-amino acids has resulted in confusion in the scientific literature and textbooks (Day & Ritter, 1967). For example the φ angles for pleated sheet structures in Table 3 of Edsall *et al.* (1966) are incorrect.

Backbones			
	<i>A</i>	<i>B</i>	
φ_1	5°	2°	
ψ_1	1	7	
ω_1	6	0	
φ_2	50	49	39° 59°
ψ_2	318	317	318 295
ω_2	4	1	0 0
φ_3	31	38	
ψ_3^1	341	337	
ψ_3^2	167	168	
Side chains			
	<i>A</i>	<i>B</i>	
N(2)–C(3)–C(14)–C(15)	(X_2^1)	307°	176°
C(4)–C(3)–C(14)–C(15)		192	62
C(3)–C(14)–C(15)–C(16)	(X_3^{22})	293	268
C(3)–C(14)–C(15)–C(17)	(X_3^{21})	174	153
N(3)–C(5)–C(7)–C(8)	(X_3^1)	54	58
C(6)–C(5)–C(7)–C(8)		295	300
C(5)–C(7)–C(8)–C(9)	(X_3^{22})	281	274
C(5)–C(7)–C(8)–C(13)	(X_3^{21})	90	84

The subscript and superscript notation used for *X* is that suggested by Ramachandran (1968).

All peptide linkages are *trans*, *i.e.* the $C'-O$ bonds are *trans* to the $N-H$ bonds, which is shown by the values for the ω angles: 6, 0, 4 and 1° . The ideal value for a planar *trans* peptide linkage is 0° .

Ramachandran, Ramakrishnan & Sasisekharan (1963) worked out two-dimensional conformational maps correlating the allowed ranges of the dihedral angles around $N-C$ and $C-C'$ in peptides containing residues with a β -carbon atom. New conventions for these angles were established by Edsall *et al.* (1966), while the subject of conformation of polypeptides and proteins has recently been reviewed (Ramachandran, 1968). This map shows small allowed areas for the right- and left-handed α -helices, and a much larger area with φ approximately between 20 and 120° , and ψ between 270 and 360° , in which the peptide chain is stretched, which is the β -structure. The conformational angles of the parallel pleated sheet structure (PPS) and anti-parallel pleated sheet structure (APPS), originally proposed by Pauling & Corey (1953) fall within the area of β -structure on the conformational map.

The φ_1 and ψ_1 values involve a glycyl residue and the φ_3 and ψ_3 angles the end-acid group. Only the φ_2 and ψ_2 angles can therefore be properly compared with standard values.

It was pointed out that in the present structure peptide dimers can be recognized. Peptide *A* is pointed in the positive *c* direction while peptide *B* is pointed in the negative *c* direction. This general feature is the basic pattern of the antiparallel pleated sheet structure. If this is the basic feature of the structure one would expect the appropriate peptide–peptide hydrogen bonds. These exist in this structure and are shown in Figs. 2 and 3: the H-bonds N(2B)–O(2A'') and O(2B)–N(2A'') are formed by the second amino acid residue. The H-bonds of the neighboring residues should then be pointed in the opposite direction and be formed by N(1), O(1), N(3) and O(3) of peptides *A* and *B*. These atoms do not form H-bonds but instead form the coordination bonds, in the expected direction, with the copper ions. One generalization of the structure therefore is that it is an APPS structure in which coordination bonds replace hydrogen bonds. The following points conform with this generalization. (a) The ψ_2 angles are very close to those calculated for the APPS model, although the φ_2 angles deviate by about 10° . (b) The translation distance between chains running in the same direction was proposed to be 9.50 Å by Pauling & Corey (1953); this compares well with the dimension of the *a* axis in this structure of 9.316 Å. (c) Pauling & Corey predicted a repeat distance of 6.68 Å along the chain; that same repeat distance in this structure is the distance between C(1) and C(5), which is 6.64 Å in molecule *A* and 6.59 Å in molecule *B*. (d) The φ_3 and ψ_3 angles are comparable with those for an APPS structure as they should be with the third residue having a β -carbon atom and the complex bonds replacing the hydrogen bonds of N(3)

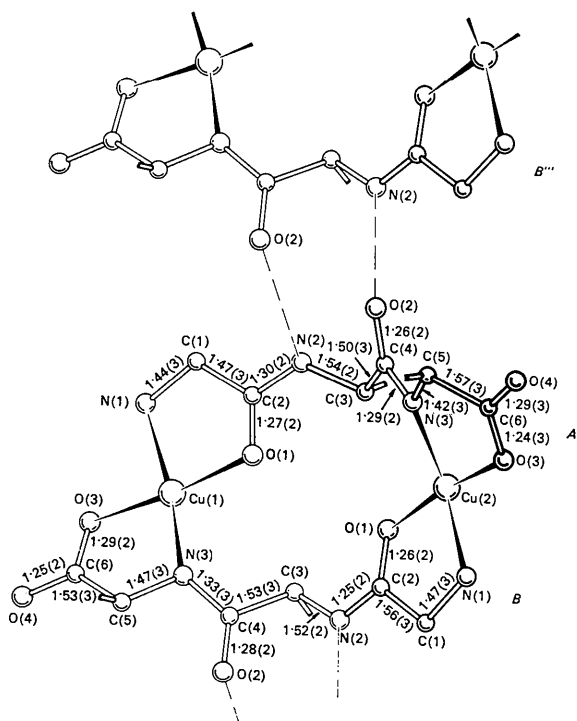


Fig. 3. Bond distances in the backbones of the peptides. The standard deviation for the last digit is given in parentheses.

and O(4). (e) The peptide itself can be crystallized (Hossain & van der Helm, 1969) and the observations obtained so far indicate that it is an APPS structure. On the basis of these observations we suggest that in general the conformational changes in peptides as a result of chelation can be expected to be small when peptide chelates are formed at neutral pH values.

The only large difference between the conformations of the two peptide units is found in the orientations of the isopropyl groups of the leucyl residues: C(14)–C(15) and N(2)–C(3) are *trans* for *B* and skewed for *A*. As noted before the C(7)–C(8) bond has to be skewed both with respect to N(3)–C(5) and C(5)–C(6) for the aromatic ring to be located below the basal plane of the square-pyramidal copper coordination. Table 8 shows that this is the case for both tyrosyl residues.

Hydrogen bonding

The hydrogen bonds involved in the formation of the sheet structure have been mentioned. The sheets are linked in the *c* direction by the complex bond O(4*B*)–Cu(2') and another peptide–peptide hydrogen bond: N(1*B*)(*b*)–O(3*B*) (Fig. 2, Table 9).

Fig. 4 shows a projection of structure down the *a* axis and is therefore a view approximately parallel to the peptide chains. The cavities between dimers are either filled by ether molecules or water molecules.

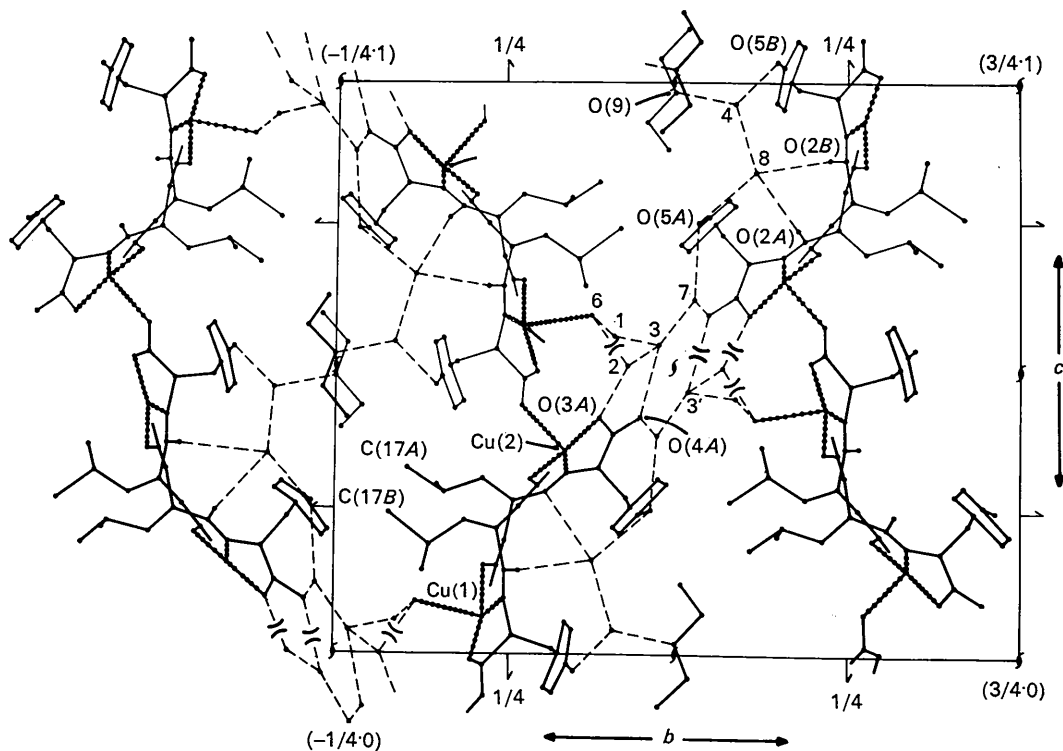


Fig. 4. Projection of the structure down the *a* axis. Atoms N(1) and C(1) for both independent peptides are not shown in this projection. The numbers indicate water molecules. Dashed lines are hydrogen bonds.

The ether molecules are stacked as a column around the 2_1 axis parallel to **a**. This column is used to fill the hole that is surrounded by groups of similar polarity, *i.e.* isopropyl groups and phenolic groups. In Fig. 4 this can be seen for instance for the column located at $(-\frac{1}{4}, \frac{1}{2})$. This explains the necessity of ether during the crystallization process. The ether oxygen atom forms an H-bond with $W(4)$. The other cavity in the structure, located at $(\frac{1}{4}, \frac{1}{2})$, is filled by water molecules.

Two hydrogen-bond spirals are present in the structure. One of these is formed by $W(7)$, $W(3)$ and the peptide oxygen atom $O(4A)$ and their symmetry equivalents by a two fold screw axis. The first spiral is connected to a second one formed by $W(6)$, $W(1)$, $W(3)$ and $W(2)$. It is doubtful if $W(1)$ - $W(3)$ is indeed

a hydrogen bond. Although the spirals are primarily used to fill the cavity, the hydrogen bond $W(2)$ - $O(3A)$ and the participation of $O(4A)$ and $W(6)$, coordinated to $Cu(1)$, indicates their packing function. The packing is also aided by a hydrogen-bond chain which extends between the ether column and the first hydrogen-bond spiral, *i.e.* $O(9)$, $W(4)$, $W(8)$, $O(5A)$ and $W(7)$. This chain has several branches: $W(4)$ - $O(5B)$, $W(8)$ - $O(2B)$ and $W(8)$ - $O(2A)$. The water molecule that has not been considered yet, $W(5)$, forms two hydrogen bonds: $W(5)$ - $O(4B)$ and $W(5)$ - $N(1A)$. The hydrogen bonds are listed in Table 9, parts A, B and C, and shown in Figs. 2 and 4. The bond not shown in the Figures is $W(1)$ - $N(1A)$, because the angle $C(1B)$ - $N(1B)$ - $W(1)$ makes it doubtful that this is indeed hydrogen bond. The alternative location of $W(1)$, *i.e.* $W(1^*)$, with

Table 9. Intermolecular distances less than 3.50 Å

The atoms in the first column have the coordinates given in Table 1. The small letters following atoms in the second column indicate the operations to be performed on the coordinates of those atoms as given in Table 1.

	<i>a</i>	$x-1,$	<i>y,</i>	<i>z</i>	
	<i>b</i>	$-x+\frac{1}{2},$	$-y,$	$z-\frac{1}{2}$	
	<i>c</i>	$-x,$	$y-\frac{1}{2},$	$-z+\frac{1}{2}$	
	<i>d</i>	$-x+1\frac{1}{2},$	$-y,$	$z+\frac{1}{2}$	
	<i>e</i>	$-x+\frac{1}{2},$	$-y,$	$z+\frac{1}{2}$	
	<i>f</i>	$x-\frac{1}{2},$	$-y-\frac{1}{2},$	$-z$	
	<i>g</i>	$-x+1,$	$y-\frac{1}{2},$	$-z+\frac{1}{2}$	
	<i>h</i>	$-x-\frac{1}{2},$	$-y,$	$z-\frac{1}{2}$	
A Peptide-peptide H-bonds					
	N(2B)-O(2A)	(<i>a</i>)	2.85 Å		
	O(2B)-N(2A)	(<i>a</i>)	2.80		
	O(3B)-N(1B)	(<i>b</i>)	2.91		
B H₂O-H₂O H-bonds					
	W(6)-W(2)		2.96 Å	W(3)-W(7)	2.75 Å
	W(8)-W(4)		2.75	W(3)-W(2)	(<i>f</i>) 2.81
	W(1)-W(3)		3.23	W(1)-W(6)	(<i>a</i>) 2.55
C H₂O-peptide and ether H-bonds					
	W(4)-O(5B)	(<i>a</i>)	2.45 Å	W(2)-O(3A)	(<i>b</i>) 2.70 Å
	W(7)-O(5A)	(<i>c</i>)	2.93	W(3)-O(4A)	(<i>b</i>) 2.87
	W(8)-O(2B)		3.06	W(7)-O(4A)	(<i>g</i>) 2.72
	W(8)-O(5A)		2.86	W(8)-O(2A)	(<i>a</i>) 2.85
	W(5)-O(4B)		2.99	W(1)-N(1A)	(<i>a</i>) 3.09
	W(5)-N(1A)	(<i>a</i>)	2.90	O(9)-W(4)	(<i>c</i>) 2.90
D W(1*) H-bonds					
	W(1*)-W(7)	(<i>f</i>)	2.96 Å		
	W(1*)-W(5)		2.98		
	W(1*)-N(1A)	(<i>a</i>)	2.87		
E All other van der Waals distances less than 3.50 Å					
	N(1B)-O(4B)	(<i>e</i>)	3.28 Å	C(17A)-O(5B)	(<i>d</i>) 3.48 Å
	W(7)-W(2)		3.46	W(4)-C(11B)	(<i>a</i>) 3.34
	W(7)-C(11A)	(<i>c</i>)	3.48	W(4)-C(10B)	(<i>a</i>) 3.33
	O(1B)-C(3A)		3.12	O(3A)-O(4B)	(<i>e</i>) 2.98
	O(1B)-O(4B)	(<i>e</i>)	3.19	W(6)-C(15B)	3.19
	C(3B)-O(1A)		3.09	W(8)-C(10A)	3.45
	C(11B)-N(1A)		3.49	W(4)-C(7B)	3.49
	N(3A)-O(4B)	(<i>e</i>)	3.29	W(5)-O(5B)	(<i>a</i>) 3.39
	O(2B)-C(1A)	(<i>a</i>)	3.16	W(5)-C(1A)	(<i>a</i>) 3.23
	O(2B)-C(2A)	(<i>a</i>)	3.43	W(5)-N(3A)	(<i>b</i>) 3.50
	C(1B)-O(2A)	(<i>a</i>)	3.17	W(1*)-W(6)	(<i>a</i>) 3.31
	C(2B)-O(2A)	(<i>a</i>)	3.45	W(1*)-N(1B)	(<i>h</i>) 3.25
	W(3)-W(2)		3.28	W(1*)-C(1B)	(<i>h</i>) 3.34
	O(2B)-O(2A)	(<i>a</i>)	3.20	W(1*)-O(4A)	(<i>b</i>) 3.26

occupancy of $\frac{1}{3}$, has better hydrogen bonds than $W(1)$ itself (Table 9, *D*). No attempt has been made to indicate donors and acceptors in the hydrogen bonding, because the hydrogen atoms were not located and several bonding schemes are feasible. The large temperature motion of the water molecules (Table 1) is probably the reason that only few reflections with $2\theta > 90^\circ$ could be observed. All other intermolecular distances less than 3.50 Å are shown in Table 9, *E*.

In summary it is interesting to note that the structure is sufficiently large and complicated for it to be possible to recognize so called hydrophilic and hydrophobic regions.

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The Crystal and Molecular Structure of 1,8-Dinitrosonaphthalene

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1,8-Dinitrosonaphthalene crystallizes in space group $P2_1/b$ with $a=13.13$, $b=16.01$, $c=3.89$ Å, $\beta=92.7^\circ$ and with $Z=4$. The structure was determined from photographic X-ray data and refined by the least-squares method to $R=0.078$ for 1009 reflexions. The molecule is in the form of an internal nitroso dimer with an N-N bond 1.38 Å long. The naphthalene nucleus shows similar distortions to those found in the acenaphthenes.

Introduction

It is possible to devise a number of plausible molecular structures for 1,8-dinitrosonaphthalene. Data from chemical and spectroscopic measurements do not allow an unambiguous distinction between these structures. Therefore the structure of the crystal was determined by X-ray diffraction.

Experimental

Red-brown crystals of the compound were supplied by Professor M. C. Whiting of Bristol University (Whiting, 1969). The crystal data are: $C_{10}H_6N_2O_2$, $M=186.2$, monoclinic,

$$a = 13.13 \pm 0.03, \quad b = 16.01 \pm 0.03, \quad c = 3.89 \pm 0.01 \text{ \AA}, \\ \alpha = 92.7 \pm 0.3^\circ, \\ Z = 4,$$

$$D_m = 1.51 \text{ g. cm}^{-3} \text{ by flotation}, \\ D_c = 1.513 \text{ g. cm}^{-3} \quad \text{Cu K}\alpha \text{ 1.5418 \AA}, \\ \mu = 9.21 \text{ cm}^{-1}.$$

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